

The glucagon-like Peptide-2

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Abstract

Multiple peptide hormones produced within the gastrointestinal system act also in the central nervous system and aid in the regulation of energy homeostasis and metabolism. The list of these peptides is progressively increasing and includes glucagon-like peptide 2 (GLP-2) as an anorexigenic factor. GLP-2 is released from enteroendocrine L-cells following food intake and its principal target is represented by the gastrointestinal tract. GLP-2 has been shown to be an important intestinotrophic factor that stimulates epithelial cell proliferation and inhibits apoptosis. GLP-2 increases intestinal blood flow and the activity and expression of epithelial brush-border digestive enzymes and nutrient transporters, and consequently increases the intestinal digestive and absorptive capacity. It inhibits gastric and intestinal motility, thus providing another mechanism to increase absorption of nutrients. Current research has focused on determining its physiological actions and its biological mechanisms in the gut, while very little is known on the GLP-2 actions within the brain. This review provides an overview of the state of the art on GLP-2 biology.

Introduction

In the last decades a wealth of data has expanded our knowledge on the glucagon-like peptide 2 (GLP-2) as important molecule involved in a wide variety of functions. GLP-2 was first discovered as an intestinotrophic factor in 1996¹, but today it is recognized as a pleiotropic hormone that—exerts different actions, not only in the gastrointestinal tract but also in the central nervous system. Since the biological actions of GLP-2 converge at multiple levels on the regulation of nutrient assimilation and energy homeostasis, a great deal of interest has been generated for drug development with therapeutic potential. The aim of this review is to explore recent advances in our understanding of GLP-2 biology.

Synthesis, secretion and degradation

GLP-2 is a 33-amino-acid peptide, which is derived from the cleavage of proglucagon, a large prohormone that is mainly expressed in cells of the endocrine pancreas, in the enteroendocrine L-cells, most of which are located in the distal ileum and colon, in the brain. Alternative splicing of proglucagon through prohormone convertases leads to the tissue-specific release of GLP-2 and other peptides with diverse biological properties. In particular, in pancreatic alpha-cells proglucagon is cleaved by prohormone convertase (PC)-2 to form glucagon, the major glucagon fragment and intervening peptide (IP)-1. In the gastrointestinal tract and in the brain, processing of proglucagon, which is operated by PC1/3, results in glucagon-like peptide-1 (GLP-1), GLP-2, IP2, oxyntomodulin and glicentin². GLP-2 is released in response to stimulation by luminal nutrient, including glucose, fatty acids and dietary fiber.

After ingestion of nutrients, plasma levels of GLP-2 increase 2- to 5-fold, depending upon the size and nutrient composition of the meal. It has been supposed that the peptide diffuse across the subepithelial lamina propria to activate afferent nerves and/or to enter the circulation, thus it may act also as paracrine agent besides as endocrine hormone³.

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GLP-2 structure is highly conserved across different mammalian species. It contains an alanine residue at position 2, rendering it ideal substrate for cleavage and inactivation by the enzyme dipeptidyl peptidase 4 (DPP-IV) and resulting in the generation of inactive GLP-2^{3,33,3}. The degradation by DPP-IV has important consequences. First of all, the half life of intravenous GLP-2 is very short, about 7 min in healthy humans⁴ and secondly, use of DPP-IV inhibitors or the substitution of alanine with glycine as in teduglutide avoids degradation and confers greater biological activity⁵.

GLP-2 receptor

GLP-2 effects are mediated by the interaction with a specific GLP-2 receptor (GLP-2R), which is a G protein-coupled receptor expressed mainly in the gut and in the brain. GLP-2R expression is restricted to the gastrointestinal tract and central nervous system, with limited expression in lung, cervix, and vagal afferents⁶⁻⁹. Multiple experimental approaches have localized the GLP-2R to regions within the rodent central nervous system (CNS) including the hippocampus, hypothalamus and nucleus of the solitary tract in the mouse¹⁰. The exact cellular localization of the GLP-2R in the gut has been a source of controversy which is probably derived from methodological problems or species-specific expression. GLP-2R may be present in enteroendocrine cells^{7,11}, enteric neurons¹²⁻¹⁴ and subepithelial myofibroblasts¹⁵, but in murine gastrointestinal tract GLP-2R is expressed exclusively in neurons and myofibroblasts, and it is not present at mucosal level^{7,12}.

In mouse, GLP-2R-mRNA has been demonstrated with high levels of expression in the bowel⁷ and recently GLP-2R protein has been demonstrated throughout the gastrointestinal tract, with higher expression in gastric fundus and colon¹⁴. The relative high prevalence of the GLP-2R in the gut might explain why, to date, GLP-2-mediated effects have been observed almost exclusively in the GI tract⁷.

Actions of GLP-2 within the gastrointestinal tract

The main biological effects of GLP-2 are related to the regulation of energy absorption and maintenance of mucosal morphology, function, and integrity¹⁶. Central to the beneficial effects of GLP-2 on the gut is its biological action to increase intestinal growth, due to the enhancement of crypt cell proliferation and inhibition of apoptosis, resulting in expansion of villous height¹. In fact, a large number of studies have demonstrated that exogenously-administered GLP-2 is tropic for the small intestine and, to a lesser extent, the colon^{1,15,17}. However, endogenous GLP-2 plays a role in the adaptative intestinal

growth that occurs in rodents in response to oral re-feeding after a period of nutrient deprivation, as shown by using GLP-2^{3,33} or GLP-2R knock out mice¹⁸⁻²⁰. The association between GLP-2 and intestinal growth/adaptation is most evident in a variety of pathological conditions, including post-resection intestinal adaptation, celiac disease, parenteral nutrition-induced intestinal atrophy and inflammatory bowel disease¹⁶.

GLP-2 is able to maintain mucosal integrity by enhancing intestinal barrier function and decreasing transcellular and paracellular epithelial permeability²¹. The ability of GLP-2 to increase barrier function has been confirmed in non-obese diabetic and ob/ob obese murine models²². In fact, administration of a prebiotic to ob/ob mice induces GLP-2-dependent up-regulation of the tight junction proteins, zonuline-1 and occludin²². In addition, GLP-2 acts in pathophysiological states as an anti-inflammatory agent, reducing intestinal mucosal inflammatory cytokine production²³.

GLP-2 exerts numerous other actions within the GI tract to promote energy absorption. It increases the uptake of luminal nutrients, including sugars and lipids, by augmenting the activity and the expression of nutrient transporters and the expression of different enzymes involved in digestion¹⁶. GLP-2 is able also to increase the mesenteric blood flow¹³. GLP-2 has also been shown to inhibit gastric acid hypersecretion²⁴ and the intestinal chloride secretion²⁵.

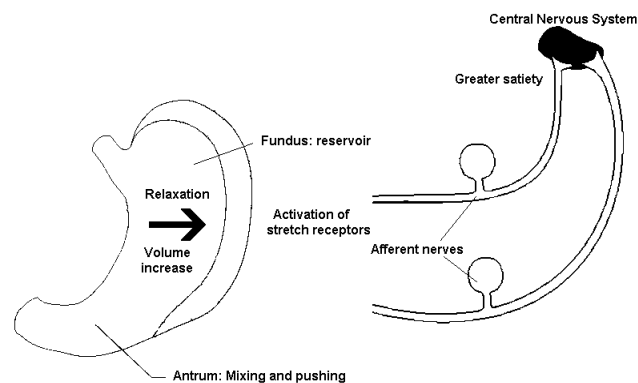


Figure 1: Increase in gastric volume induced by GLP-2 might represent satiety signaling to the central nervous system through the activation of stretch receptors and afferent nerves

GLP-2 inhibits GI motility, thus providing another mechanism to increase digestion and absorption of

nutrient. Specifically, GLP-2 reduces the vagally-induced antral motility in pigs²⁶ and it decreases the mouse gastric fundic tone leading to an increase of stomach capacity²⁷. Although it is not clearly established if the GLP-2 effect on mouse gastric stomach is physiological or pharmacological, the GLP-2 action on gastric fundus seems particularly interesting, because it could represent a satiety signaling which well fits with the finding that GLP-2 is a chemical mediator inhibiting rodent feeding behaviour²⁸. In fact, increase in the gastric volume might mean activation of stretch receptors and greater satiety signals to the brain (Fig.1), but electrophysiological experiments *in vivo* are necessary to confirm this hypothesis.

In mouse GLP-2 inhibits the intestinal transit *in vivo*²⁹ and it reduces spontaneous or electrically-evoked cholinergic contractions of the small and large intestine *in vitro*^{14,30}. The peptide modulation on the gastrointestinal motility may be due to central nervous mechanisms²⁶, but involvement of the enteric nervous system has been also clearly shown through the *in vitro* studies^{14,27,30}. As GLP-2R is expressed in the subepithelial myofibroblasts and enteric nervous system as well as human enteroendocrine cells, it has been proposed that the peptide exerts its actions indirectly via downstream mediators deriving from GLP-2R-expressing cells⁷. Indeed, neural VIP, nitric oxide and reduction of the acetylcholine release from enteric nerves have been reported to be involved in the inhibitory motor effects induced by GLP-2 in different regions of the mouse GI tract^{14,27,30}. However determining how GLP-2 produces its biological effects, which mediators are involved and how these mediators interact is an area of intense research³¹ and the reader is referred to a recent paper for a more detailed overview³².

Extraintestinal actions of GLP-2

GLP-2 may be considered an anorexigenic peptide because intracerebroventricular administrations of GLP-2 reduce the food intake in rodents^{8,28}. Interestingly, in rats the anorectic response to GLP-2 seems to depend on a certain tone of central GLP-1 receptors because pharmacological antagonism of GLP-1 receptors by prior administration of exendin(9–39) abolishes GLP-2 induced anorexia²⁸. On the contrary studies in mice have pointed to the opposite: Blocking central GLP-1 receptors with exendin-9 increases GLP-2 induced anorexia⁸. Further studies focusing on the role of central GLP-2 receptors in appetite regulation are clearly needed. On the other hand, multiple experimental approaches have localized the GLP-2R to regions within the rodent CNS including the hypothalamus and hippocampus¹⁰. Activation of the GLP-2R in the hypothalamus may be responsible for the effects of central GLP-2 administration on feeding behaviour and

because the hippocampus is sensitive to hunger and satiety signals, might influence appetitive behaviour through inhibitory learning and memory process. However, up to date studies in humans have not demonstrated decrease in the food intake after peripheral GLP-2 administration^{33,34}, even if recent data have shown that intraperitoneal injections of teduglutide reduce food intake in mouse, suggesting a role for GLP-2 in the short-term regulation of the ingestive behaviour³⁵.

In general, very few studies have been conducted to elucidate the roles of the GLP-2 in the central nervous system³⁶. Consistent with a general cytoprotective effect of GLP-2 within the GI mucosa, the few studies have suggested that activation of GLP-2 receptors can protect neurons from excitotoxic damage³⁶. More specifically, GLP-2 has been reported to reduce glutamate-induced cell death in cultured hippocampal cells¹⁰ and to stimulate the proliferation of rat astrocytes^{37,38}. Antidepressant-like effects of GLP-2 that occur via monoamine pathways have also been noted in mice, but this has yet to be confirmed³⁹.

Conclusions and future directions

All the actions described make GLP-2 an attractive drug for the treatment or prevention of gastrointestinal disease in human subjects. However, because very few studies have been conducted to elucidate the roles of this peptide in central nervous system, we are still far from having an in depth understanding of the complex neurobiology of GLP-2. Therefore, future research efforts should aim at providing new information on the roles subserved by GLP-2 within the brain.

References

1. Drucker DJ, Erlich P, Asa SL, Brubaker PL. Induction of intestinal epithelial proliferation by glucagon-like peptide 2. *Proc Natl Acad Sci.* 1996;93:7911-6.
2. Wallis K, Walters JRF, Forbes A. Review article: glucagon-like peptide-2-current applications and future directions. *Aliment Pharmacol Ther.* 2007;25:365-72.
3. Cummings DE, Overduin J. Gastrointestinal regulation of food intake. *J Clin Invest.* 2007;117:13-23.
4. Hartmann B, Harr MB, Jeppesen PB, Wojdemann M, Deacon CF, Mortensen PB, et al. *In vivo* and *in vitro* degradation of glucagon like peptide-2 in humans. *J Clin Endocrinol Metab.* 2000;85:2884-8.
5. Jeppesen PB, Sanguinetti EL, Buchman A, Howard L, Scolapio JS, Ziegler TR, et al. Teduglutide(ALX-0600), a dipeptidylpeptidase IV resistant glucagon-like peptide 2 analogue, improves intestinal function in short bowel syndrome patients. *Gut* 2005;54:1224-31.
6. Munroe DG, Gupta AK, Kooshesh P, Vyas TB, Rizkalla G, Wang H, et al. Prototypic G protein-coupled receptor for the intestinotrophic factor glucagon-like peptide 2. *Proc Natl Acad Sci.* 1999;96:1569-73.

7. Yusta B, Huang L, Munroe D, Wolff G, Fantáske R, Sharma S, et al. Enteroendocrine localization of GLP-2 receptor expression in humans and rodents. *Gastroenterology* 2000;119: 744-55.
8. Lovshin J, Estall J, Yusta B, Brown TJ, Drucker J. Glucagon-like peptide (GLP)-2 action in the murine central nervous system is enhanced by elimination of GLP-1 receptor signaling. *J Biol Chem* 2001;276:21489-99.
9. Nelson DW, Sharp JW, Brownfield MS, Raybould HE, Ney DM. Localization and activation of glucagons-like peptide-2 receptors on vagal afferents in the rat. *Endocrinology* 2007;148:1954-62.
10. Lovshin JA, Huang Q, Seaberg R, Brubaker PL, Drucker DJ. Extrahypothalamic expression of the glucagon-like peptide-2 receptor is coupled to reduction of glutamate-induced cell death in culture hippocampal cells. *Endocrinology* 2004;145:3495-506.
11. Guan X, Stoll B, Lu X, Tappenden KA, Holst JJ, Hartmann B, et al. GLP-2-mediated up-regulation of intestinal blood flow and glucose uptake is nitric oxide-dependent in TPN-fed piglets. *Gastroenterology* 2003;125:136-47.
12. Bjerknes M, Cheng H. Modulation of specific intestinal epithelial progenitors by enteric neurons. *Proc Natl Acad Sci*. 2001;98:12497-502.
13. Guan X, Karpen HE, Stephens J, Bukowski JT, Niu S, Zhang G, et al. GLP-2 receptor localised to enteric neurons and endocrine cells expressing vasoactive peptides and mediates increased blood flow. *Gastroenterology* 2006; 130:150-64.
14. Amato A, Rotondo A, Cinci L, Baldassano S, Vannucchi MG, Mulè F. Role of cholinergic neurons in the motor effects of glucagon-like peptide-2 in mouse colon. *Am J Physiol Gastrointest Liver Physiol*. 2010;299:G1038-44.
15. Ørskov C, Hartmann B, Poulsen SS, Thulesen J, Hare KJ, Holst JJ. GLP-2 stimulates colonic growth via KGF, released by subepithelial myofibroblasts with GLP-2 receptors. *Regul Pept*. 2005;124:105-12.
16. Estall JL, Drucker DJ. Glucagon-like peptide-2. *Annu Rev Nutr*. 2006;26:391-411.
17. Dubé PE, Forse CL, Bahrami J, Brubaker PL. The essential role of insulin-like growth factor-1 in the intestinal tropic effects of glucagon-like peptide-2 in mice. *Gastroenterology* 2006;131:589-605.
18. Shin ED, Estall JL, Izzo A, Drucker DJ, Brubaker PL. Mucosal adaptation to enteral nutrients is dependent on the physiologic actions of glucagon-like peptide-2 in mice. *Gastroenterology* 2005;128:1340-53.
19. Nelson DW, Murali SG, Liu X, Koopmann MC, Holst JJ, Ney DM. Insulin-like growth factor I and glucagon-like peptide-2 responses to fasting followed by controlled or ad libitum refeeding in rats. *Am J Physiol Regul Integr Comp Physiol*. 2008;294:R1175-84.
20. Bahrami J, Yusta B, Drucker DJ. ErbB activity links the glucagon-like peptide-2 receptor to refeeding-induced adaptation in the murine small bowel. *Gastroenterology* 2010;138:2447-56.
21. Benjamin MA, McKay DM, Yang PC, Cameron H, Perdue MH. Glucagon-like peptide-2 enhances intestinal epithelial barrier function of both transcellular and paracellular pathways in the mouse. *Gut* 2000;47:112-9.
22. Cani PD, Possemiers S, Van de Wiele T, Guiot Y, Everard A, Rottier O, et al. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. *Gut* 2009;58:1091-103.
23. Sigalet DL, Wallace LE, Holst JJ, Martin GR, Kaji T, Tanaka H, et al. Enteric neural pathways mediate the anti-inflammatory actions of glucagon-like peptide 2. *Am J Physiol Gastrointest Liver Physiol*. 2007;293:G211-21.
24. Wøjdemann M, Wettergren A, Hartmann B, Hilsted L, Holst JJ. Inhibition of sham feeding-stimulated human gastric acid secretion by glucagon-like peptide-2. *J Clin Endocrinol Metab*. 1999;84:2513-7.
25. Baldassano S, Liu S, Qu MH, Mulè F, Wood JD. Glucagon-like peptide-2 modulates neurally evoked mucosal chloride secretion in guinea pig small intestine in vitro. *Am J Physiol Gastrointest Liver Physiol*. 2009; 297:G800-5.
26. Wøjdemann M, Wettergren A, Hartmann B, Holst JJ. Glucagon-like peptide-2 inhibits centrally induced antral motility in pigs. *Scand J Gastroenterol*. 1998;33:828-32.
27. Amato A, Baldassano S, Serio R, Mulè F. Glucagon-like peptide-2 relaxes mouse stomach through vasoactive intestinal peptide release. *Am J Physiol Gastrointest Liver Physiol*. 2009;296:G678-84.
28. Tang-Christensen M, Larsen PJ, Thulesen J, Romer J, Vrang N. The proglucagon-derived peptide, glucagon-like peptide-2, is a neurotransmitter involved in the regulation of food intake. *Nat Med* 2000;6:802-7.
29. McDonagh SC, Lee J, Izzo A, Brubaker PL. Role of glial cell-line derived neurotropic factor family receptor $\alpha 2$ in the actions of the glucagon-like peptides on the murine intestine. *Am J Physiol Gastrointest Liver Physiol*. 2007;293: G461-G468.
30. Cinci L, Fausson-Pellegrini MS, Rotondo A, Mulè F, Vannucchi MG. GLP-2 receptor expression in excitatory and inhibitory enteric neurons and its role in mouse duodenum contractility. *Neurogastroenterol Motil*. 2011;doi: 10.1111/j.1365-2982.2011.01750.x
31. Dubé PE, Brubaker PL. Frontiers in glucagon-like peptide-2: multiple actions, multiple mediators. *Am J Physiol Endocrinol Metab*. 2007;293:E460-5.
32. Rowland KJ, Brubaker PL. The "cryptic" mechanism of action of glucagon-like peptide-2. *Am J Physiol Gastrointest Liver Physiol*. 2011;301:G1-8.
33. Schmidt PT, Hartmann B, Bregenholt S, Hoist JJ, Claesson MH. Deficiency of the intestinal growth factor, glucagon-like peptide 2, in the colon of SCID mice with inflammatory bowel disease induced by transplantation of CD4+ T cells. *Scand J Gastroenterol*. 2000;35:522-7.
34. Sørensen LB, Flint A, Raben A, Hartmann B, Holst JJ, Astrup A. No effect of physiological concentrations of glucagon-like peptide-2 on appetite and energy intake in normal weight subjects. *Int J Obes Relat Metab Disord*. 2003;27:450-6.
35. Baldassano S, Mulè F. 2011. Peripheral glucagons like peptide 2 analogue administration reduces food intake in lean and diet-induced obese mice. *Gastroenterology* 140(5) suppl. 1 s134.
36. Vrang N, Larsen PJ. Preproglucagon derived peptides GLP-1, GLP-2 and oxyntomodulin in the CNS: role of peripherally secreted and centrally produced peptides. *Prog Neurobiol*. 2010;92:442-62.
37. Velázquez E, Ruiz-Albusac JM, Blázquez E. Glucagon-like peptide-2 stimulates the proliferation of cultured rat astrocytes. *Eur J Biochem*. 2003;270:3001-9.
38. Velázquez E, Blázquez E, Ruiz-Albusac JM. Synergistic effect of glucagon-like peptide 2 (GLP-2) and of key growth factors on the proliferation of cultured rat astrocytes. Evidence for reciprocal upregulation of the mRNAs for GLP-2 and IGF-I receptors. *Mol Neurobiol*. 2009;40:183-93.
39. Iwai T, Hayashi Y, Narita S, Kasuya Y, Jin K, Tsugane M, et al. Antidepressant-like effects of glucagon-like peptide-2 in mice occur via monoamine pathways. *Behav Brain Res*. 2009;204:235-40.

