



Effect of the 5-HT_{2C} Receptor Agonist WAY-163909 on Serotonin and Dopamine Metabolism across the Rat Brain: A Quantitative and Qualitative Neurochemical Study

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Abstract: The effects triggered by serotonin2C (5-hydroxytryptamin_{2C}, 5-HT_{2C}) receptor agonists in the brain are often subtle, and methodologies highlighting their widespread actions to account for their multiple modulatory influences on behaviors are still lacking. We report an extended analysis of a neurochemical database on monoamines obtained after the intraperitoneal administration of the preferential 5-HT_{2C} receptor agonist WAY-163909 (0.3 and 3 mg/kg) in 29 distinct rat brain regions. We focused on the metabolite of 5-HT, 5-hydroxyindoleacetic acid (5-HIAA), the metabolites of dopamine (DA), 3,4-dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA), and the index of the turnovers 5-HIAA/5-HT and DOPAC/DA. WAY-163909 increased and decreased 5-HIAA tissue levels in the amygdala and dorsolateral orbitofrontal cortex, respectively, and decreased the 5-HT turnover in the infralimbic cortex. It enhanced HVA levels in the medial orbitofrontal cortex and DOPAC levels in the amygdala. WAY-163909 increased and decreased DA turnover in the medial orbitofrontal cortex and the anterior insular cortex, respectively. The correlative analysis of the turnovers between pairs of brain regions revealed low levels of correlations across the brain but presented a distinct pattern of correlations after WAY-163909 was compared to saline-treated rats. WAY-163909, notably at 0.3 mg/kg, favored cortico-cortical and cortico-subcortical correlations of both turnovers separately, and frontal DOPAC/DA ratio with cortical and subcortical 5-HIAA/5-HT ratios at 3 mg/kg. In conclusion, the qualitative, but not the quantitative analysis shows that WAY-163909 alters the pattern of correlations across the brain, which could account for its multiple behavioral influences.

Keywords: metabolism; dopamine; serotonin; 5-HIAA; HPLC; HVA; DOPAC; correlative analysis; 5-HT_{2C} receptor; connectivity



1. Introduction

The serotonin2C (5-hydroxytryptamin_{2C}, 5-HT_{2C}) receptor, a seven-transmembrane receptor coupled to several intracellular signaling pathways [1], is thought to act as a "neural rheostat regulating the intersection between vulnerability behaviors" [2]. This hypothesis parallels the initial proposal that 5-HT_{2C} receptors regulate or even inhibit the excitability of neurobiological networks [3]. The ability of 5-HT_{2C} receptors to shape neurobiological networks is widespread, and the receptor has been shown to interfere with both the serotonin (5-HT) and dopamine (DA) systems in the mammalian brain. The stimulation of 5-HT_{2C} receptors has been shown in rodents to reduce feeding behavior [4,5], drug intake [6–8], impulsive behavior [9,10], or psychosis [11] and to trigger anxiety [12–14] and compulsive forms of behaviors [15,16], to cite a few [1]. It could also participate in the mechanism of action of some hallucinogenic drugs, including d-Lysergic acid diethylamide [17]. The understanding of 5-HT_{2C} receptor function is critical for the development of therapeutic drugs for the treatment of depression, schizophrenia, and drug abuse [1,18], but it presents a major neurobiological challenge.

The widespread expression of the 5-HT_{2C} receptor subtype in the CNS [19–22], and the paucity of the responses to 5-HT_{2C} receptor agonists obtained with classical neuroanatomical markers of neuronal activity, such as c-Fos [23–25], makes it difficult to study the consequences of its stimulation on neurobiological networks. There are numerous data reporting that the stimulation of 5-HT_{2C} receptors alters the activity of neuronal cells in brain tissues *in vitro* and *in vivo* [1,23]. However, it is also difficult to understand the relationship between these neuronal responses to 5-HT_{2C} receptor stimulation, their consequences on a neurobiological network and, ultimately, the behaviors. The ascending 5-HT systems of the dorsal and median raphe nuclei (DR and MR) innervate the brain quite diffusely, while the ascending DA systems of the substantia nigra (SN) and ventral tegmental area (VTA) have a smaller number of prescribed circuits [26]. The biochemical activity at their terminal fields includes both homologous and heterologous controls, the latter involving neighboring cells interacting with neurobiological networks [27,28]. The neurochemical approach of analyzing tissue monoamine content across several parts of the brain is a possibility to study extrinsic influences operating on terminals of monoaminergic neurons.

Recently, we reported that the preferential agonist WAY-163909 modified the tissue content of 5-HT and DA only in a few brain regions. However, it reduced the correlations of DA tissue content established between pairs of brain regions and redistributed the pattern of correlations for DA and 5-HT content across the brain [29]. The measurement of metabolites of monoamines can inform other aspects of monoamine neuron function compared to the neurotransmitter itself [30]. Previous studies have shown clear regional differences in basal neurotransmitter release, metabolite levels, and metabolite turnover rates [31–33]. 5-hydroxyindoleacetic acid (5-HIAA) and 3,4-dihydroxyphenylacetic acid (DOPAC) come from the breakdown of 5-HT and DA, respectively, via a first oxidation by monoamine oxidase A (MAO-A) followed by aldehyde dehydrogenase [34]. Homovanillic acid (HVA) is the final product of the degradation of DA involving the catechol-O-methyl transferase (COMT) [27,34]. In contrast to the neurotransmitter tissue content, mainly corresponding to compounds contained in vesicles of exocytosis at terminals [27], metabolites also come from the activity of surrounding cells. DOPAC can be produced both inside and outside catecholaminergic cells. For 5-HIAA, MAO-A is not often present at terminals of 5-HT neurons; therefore, this process likely occurs outside the terminals in the case of 5-HT [27,35–37]. Similarly, COMT is located elsewhere other than catecholaminergic cells [38]. Meanwhile, the 5-HIAA/5-HT and DOPAC/DA ratios, which are considered as indirect indexes of the turnover, vary across the brain, and the pattern of correlations between pairs of specific brain regions is different for the parent neurotransmitter [32,39]. It is, therefore, possible that the stimulation of central 5-HT_{2C} receptors alters the metabolism of 5-HT and DA in a different way compared to the neurotransmitter itself.

In the present study, we have expanded the analysis of the effect of the selective 5-HT_{2C} receptor agonist WAY-163909 (0.3 and 3 mg/kg, intraperitoneally (i.p.)) on 5-HT and DA function by studying its quantitative and qualitative neurochemical impact on metabolites of 5-HT and DA, as well as

their turnovers. The analysis was done across 29 brain regions, including the cortex, basal ganglia, mesencephalon, amygdala, hippocampus, and hypothalamus.

2. Results

2.1. Quantitative Analysis of DA and 5-HT Metabolism after WAY-163909 Treatment

We studied the effect of WAY-163909 treatment (0.3 and 3 mg/kg i.p.) on DA and 5-HT metabolism in 29 brain areas of rats and compared it with saline-treated rats (8 rats in each of the three groups; n = 24 for the entire experiment) (Figure 1). This postmortem analysis corresponds to 45 minutes after the injection of WAY-163909 (see Methods). The quantitative distribution of tissue 5-HIAA paralleled that previously reported for 5-HT [29]. The highest 5-HIAA level was found in the SN (738 ± 55.9 pg/mg) (Table 1). The ratio of 5-HIAA/5-HT was also heterogeneous in brain ranging from low (dorsomedial striatum (DMS), dorsolateral striatum (DLS), pins, ains) to high in the MR, dorsal hippocampus (dHP), and lateral orbitofrontal cortex (LO) (Figure 1A).

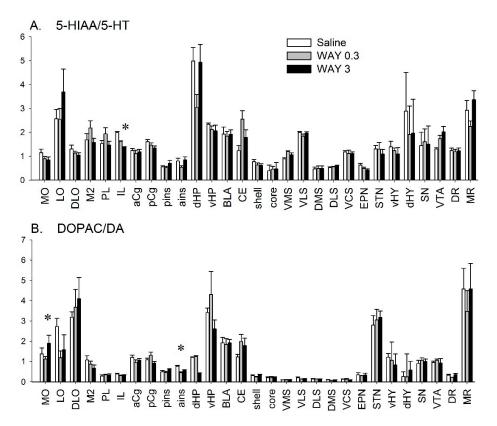


Figure 1. Effect of WAY-163909 on turnover of 5-HT (5-HIAA/5-HT ratio) (**A**) and DA (DOPAC/DA ratio) (**B**) across rat brain regions. The left, medial, and right panels correspond to saline-, WAY-163909 0.3 mg/kg, and 3 mg/kg treated rats, respectively. The results correspond to the mean \pm SEM of the ratios in 29 different brain regions. WAY-163909 or saline has been intraperitoneally (i.p.) administered, and the tissue values correspond to 45 minutes after the administration. Data for DA and 5-HT tissue contents used in this experiment have been already reported [29]. WAY-163909's effects have been compared to saline-treated rats using a one-way ANOVA. The number of values considered has been reported in Table S1. * *p* < 0.05 with respect to the saline treatment group (protected least significant difference (PLSD) test).

WAY-163909 (0.3 mg/kg) reduced 5-HIAA levels in the dorsolateral orbitofrontal cortex (DLO) and increased them in the central nucleus of the amygdala (CE) (F(2,23) = 3.61, p < 0.05, and F(2,22) = 5.307, p < 0.05, respectively) (Table 1). WAY-163909 decreased the ratio of 5-HIAA/5-HT in the infralimbic cortex (IL) (F(2,23) = 3.694, p < 0.05) at 3 mg/kg only (Figure 1A).

| Brain Regions | 5-HIAA | | | DOPAC | | | HVA | | |
|---------------|-----------------|-----------------|-----------------|-----------------|-----------------|------------------|-----------------|-----------------|-----------------|
| | Saline | WAY 0.3 | WAY 3 | Saline | WAY 0.3 | WAY 3 | Saline | WAY 0.3 | WAY 3 |
| МО | 206 ± 30 | 224 ± 20 | 252 ± 19 | 122 ± 22 | 121 ± 15 | 180 ± 22 | 23.9 ± 3.12 | 91.4 ± 20 * | 103 ± 24 ** |
| LO | 212 ± 34 | 188 ± 26 | 328 ± 54 | 35.8 ± 2.81 | 44.1 ± 15.3 | 66.1 ± 9.22 | nd | nd | nd |
| DLO | 73.6 ± 11.9 | 37.1 ± 2.95 * | 59.1 ± 11.3 | 13.2 ± 2.88 | 12.6 ± 1.8 | 20.3 ± 3.92 | nd | nd | nd |
| M2 | 92.2 ± 20.1 | 87.3 ± 12.6 | 104 ± 15.6 | 27.6 ± 3.09 | 20.9 ± 1.7 | 30.4 ± 5.76 | 1.82 ± 0.61 | 1.28 ± 0.34 | 1.99 ± 0.35 |
| PL | 137 ± 10.9 | 149 ± 24.7 | 116 ± 13.9 | 9.12 ± 1.30 | 12.5 ± 3.43 | 9.71 ± 1.95 | 36.2 ± 11.7 | 103 ± 20.3 | 62.9 ± 19.7 |
| IL | 147 ± 35.3 | 124 ± 19.1 | 121 ± 31.2 | 29.9 ± 5.16 | 19.4 ± 1.94 | 25.5 ± 6.39 | 18.5 ± 3.92 | 16.3 ± 3.21 | 24.6 ± 4.94 |
| aCg | 336 ± 16.0 | 312 ± 72.6 | 228 ± 45.1 | 180 ± 14.1 | 118 ± 34.6 | 115 ± 27.5 | nd | nd | nd |
| pCg | 386 ± 43.8 | 348 ± 34.5 | 308 ± 16.8 | 41.9 ± 3.28 | 38.0 ± 4.49 | 30.3 ± 1.46 | nd | nd | nd |
| ains | 136 ± 18.9 | 113 ± 20 | 106 ± 11.8 | 24.5 ± 5.13 | 27.9 ± 4.72 | 21.2 ± 2.4 | 8.86 ± 2.75 | 6.67 ± 1.22 | 6.15 ± 0.71 |
| pins | 99.6 ± 8.35 | 79 ± 11.7 | 102 ± 15.9 | 17.1 ± 4.1 | 15.2 ± 1.6 | 16.9 ± 3.63 | 3.98 ± 0.56 | 3.75 ± 0.76 | 4.22 ± 0.70 |
| dHP | 81.8 ± 9.15 | 60.3 ± 8.36 | 81.8 ± 4.75 | 12.3 ± 0.34 | 14.24 ± 0.90 | 12.43 ± 0.67 | nd | nd | nd |
| vHP | 159 ± 3.61 | 139 ± 17.6 | 132 ± 11.9 | 15.2 ± 2.2 | 15.5 ± 3.04 | 13.71 ± 4.2 | nd | nd | nd |
| BLA | 151 ± 17.7 | 215 ± 25.9 | 229 ± 17.5 | 104 ± 53.9 | 116 ± 23.6 | 100 ± 32.7 | nd | nd | nd |
| CE | 132 ± 18.6 | 199 ± 21.4 * | 108 ± 19.9 | 59.0 ± 10.0 | 131 ± 40.3 | 45.8 ± 13.9 | nd | nd | nd |
| shell | 178 ± 17.5 | 123 ± 10.5 | 154 ± 16.1 | 728 ± 85.8 | 595 ± 73.0 | 726 ± 71.0 | 540 ± 83.6 | 449 ± 41.4 | 586 ± 52.8 |
| core | 76.0 ± 6.36 | 63.8 ± 7.52 | 73.0 ± 11.1 | 847 ± 78 | 682 ± 106 | 825 ± 145 | 150 ± 9.02 | 124 ± 16.1 | 166 ± 23.5 |
| VMS | 142 ± 23.8 | 137 ± 13.8 | 170 ± 23.5 | 480 ± 80.2 | 477 ± 72.6 | 569 ± 84.6 | 257 ± 56.7 | 272 ± 48.4 | 352 ± 107 |
| VLS | 208 ± 12.3 | 244 ± 18.2 | 274 ± 28.2 | 876 ± 123 | 661 ± 49.6 | 799 ± 58.6 | 407 ± 62.8 | 363 ± 15.2 | 445 ± 36.6 |
| DMS | 96.2 ± 15.9 | 85.6 ± 6.45 | 79.1 ± 7.23 | 709 ± 109 | 709 ± 98.6 | 720 ± 83.1 | 603 ± 95.8 | 667 ± 66.4 | 806 ± 59.0 |
| DLS | 190 ± 66.9 | 110 ± 17.6 | 144 ± 20.3 | 797 ± 129 | 727 ± 92.5 | 838 ± 110 | 982 ± 93.3 | 827 ± 97.0 | 1113 ± 167 |
| VCS | 172 ± 36.9 | 173 ± 38.4 | 250 ± 42.2 | 307 ± 65.4 | 272 ± 76.9 | 471 ± 82.6 | 359 ± 80.6 | 359 ± 96.8 | 521 ± 94.2 |
| EPN | 147 ± 17.2 | 174 ± 25.8 | 111 ± 15.4 | 16.7 ± 2.92 | 18.6 ± 3.81 | 11.40 ± 2.11 | 1.76 ± 0.27 | 2.38 ± 0.65 | 1.79 ± 0.23 |
| STN | 257 ± 43.4 | 260 ± 37.6 | 310 ± 34.9 | 132 ± 32.5 | 148 ± 22.6 | 181 ± 29.7 | 6.48 ± 2.15 | 7.57 ± 1.29 | 9.05 ± 1.40 |
| vHY | 74.9 ± 8.82 | 54.3 ± 6.15 | 51.8 ± 8.67 | 28.2 ± 7.14 | 22.8 ± 4.02 | 25.0 ± 3.40 | nd | nd | nd |
| dHY | 156 ± 23.6 | 101 ± 14.9 | 81 ± 21.6 | 3.16 ± 0.62 | 2.19 ± 0.52 | 2.41 ± 0.63 | 2.63 ± 0.77 | 1.86 ± 0.57 | 3.87 ± 1.29 |
| SN | 738 ± 55.9 | 766 ± 47.9 | 754 ± 37.8 | 474 ± 82.4 | 436 ± 70.1 | 642 ± 61.7 | 194 ± 59.8 | 133 ± 34.5 | 176 ± 50.9 |
| VTA | 428 ± 44.3 | 459 ± 41.4 | 385 ± 62.8 | 625 ± 64.3 | 650 ± 91.4 | 597 ± 67.6 | 83.0 ± 10.8 | 85.0 ± 8.17 | 75.4 ± 10.5 |
| DR | 396 ± 32.6 | 373 ± 19.8 | 407 ± 33.1 | 23.2 ± 5.9 | 19.2 ± 3.9 | 31.9 ± 7.3 | nd | nd | nd |
| MR | 318 ± 62 | 230 ± 35 | 265 ± 40.5 | 63.9 ± 18.8 | 107 ± 28.8 | 37.9 ± 19.2 | 15.4 ± 1.8 | 16.4 ± 2.9 | 9.13 ± 1.7 |

Table 1. Tissue content of 5-HIAA, DOPAC, and homovanillic acid (HVA) in brain regions of saline and WAY-163909-treated rats.

Data for 5-HIAA, DOPAC, and HVA are mean \pm SEM values (pg/mg of tissue) per group in the 29 brain regions. The number of values retained in each group in each group has been reported in Table S1. It usually comprises between 6–8 values/group with some exceptions (notably: DOPAC in LO for saline-treated rats, n = 4; DOPAC in dHP for 3 mg/kg, n = 3). * p < 0.05, ** p < 0.01 (PLSD test) compared to saline-treated rats. nd—not detected.

The pattern of quantitative distribution of the two main metabolites of DA, namely, DOPAC and HVA, is similar to the parent neurotransmitter [29], very concentrated in the striatum and NAc, substantially concentrated in the SN or VTA, and poorly present in other brain regions including the various cortical, hippocampal, and hypothalamic subregions (Table 1). The levels of HVA were also below the threshold of detection in several brain regions, in part due to the lower electrochemical sensitivity of our apparatus for HVA compared to DOPAC or DA (see methods). The DOPAC/DA ratio was heterogeneous between the structures of the brain. It was low in the striatum (close to 0.1 and 0.15 or 0.2 in the nucleus accumbens) and high in some cortical areas, the subthalamic nucleus (STN), the hippocampus or the MR (Figure 1B).

WAY-163909 did not alter DOPAC levels in any regions whereas it increased HVA tissue content in MO (F(2,23) = 3.61, p < 0.05) only (Table 1). The effect was observed for both doses of WAY-163909 with a similar magnitude. WAY-163909 modified the DA turnover in the MO (F(2, 22) = 3.53, p < 0.05) and ains (F(2,18) = 12.2, p < 0.001). Specifically, the DA turnover was increased in MO at a dose of 3 mg/kg and decreased in ains at a dose of 0.3 mg/kg (Figure 1B).

2.1.1. Pattern of Correlations of the 5-HIAA/5-HT Ratio across the Brain

We evaluated the effect of WAY-163909 on 5-HT metabolism by studying the correlations of the 5-HIAA/5-HT ratio between pairs of regions across the 29 sampled brain regions (Figure 2). The correlations have been corrected as previously reported (see Methods). In saline-treated rats, we reported 38 significant correlations for the 5-HIAA/5-HT ratio equilibrated between positive (29) and negative (9) correlations. This represents about 9% of pairs of the entire analysis. Regions such as STN or ains established several correlations with other brain regions. Interestingly, the number of correlations remained almost constant after WAY-163909 (35 comprising 21 positive and 14 negative correlations after 0.3 mg/kg; 41 comprising 21 positive and 20 negative correlations after 3 mg/kg), but the pattern of correlations was different among the treatment groups. WAY-163909 enhanced the correlations for the 5-HT turnover in IL, aCg, shell, and entodepuncular nucleus (EPN) at 0.3 mg/kg compared to saline-treated rats. By contrast, it enhanced the correlations in LO, M2, and MR at 3 mg/kg. These changes of pattern induced by WAY-163909 occurred at the expense of the correlations in the STN, as well as the CE and SN. WAY-163909 also changed the pattern of correlations found in the dHY at 0.3 mg/kg compared to saline-treated rats, with correlations centered no longer on the dHY but the ventral hypothalamus (vHY) instead at 3 mg/kg. To summarize, from a heterogeneous distribution in saline-treated rats, WAY-163909 favored correlations between cortical and subcortical areas at 0.3 mg/kg, but not at 3 mg/kg (Figure 3).

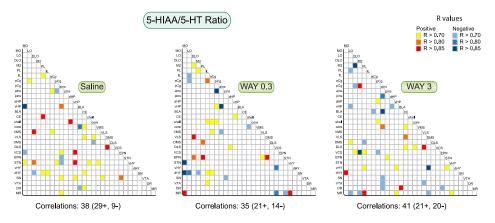


Figure 2. Correlative analysis of neurochemical indexes of 5-HT turnover (5-HIAA/5-HT ratio) across rat brain regions. Representation of the range of Pearson's *r* values for each linear regression, 5-HIAA/5-HT ratio between the 29 brain areas in saline, WAY-163909 0.3 mg/kg, and 3 mg/kg treated rats. Colored boxes correspond to the existence of a significant correlation between the two parameters (yellow to red: positive; blue: negative) considered after correction for multiple comparisons.

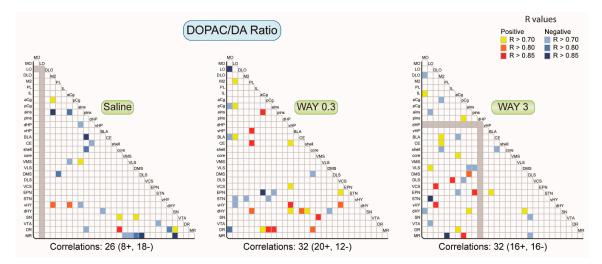


Figure 3. Correlative analysis of neurochemical indexes of DA turnover (DOPAC/DA ratio) across rat brain regions. Representation of the range of Pearson's *r* values for each linear regression, DOPAC/DA ratio between the 29 brain areas in saline, WAY-163909 0.3 mg/kg, and 3 mg/kg treated rats. Colored boxes correspond to the existence of a significant correlation between the two parameters (yellow to red: positive; blue: negative) considered after correction for multiple comparisons. The grey boxes indicate that these correlations were not performed due to the low number of values (see Table S1).

2.1.2. Pattern of Correlations of the DOPAC/DA Ratio across the Brain

We evaluated the effect of WAY-163909 on the DOPAC/DA ratio. The saline-treated rats showed 26 correlations for the DOPAC/DA ratio comprising 8 positive and 18 negative correlations (Figure 3). The distribution did not reveal a specific pattern or organization, except the slightly higher number of correlations established with the MR, and did not show the expected correlations between the quadrants of the striatum. WAY-163909 slightly increased the number of correlations to 32 at both 0.3 and 3 mg/kg. Beyond the balance of positive/negative correlations, which was different between the two doses (20 positive and 12 negative correlations at 0.3 mg/kg versus 16 positive and 16 negative correlations at 3 mg/kg), the pattern of correlations also differed. WAY-163909 (0.3 mg/kg) suppressed the number of correlations as compared to saline-treated rats in the M2, SN, and MR. Conversely, WAY-163909 potentiated the correlations in the orbitofrontal cortex (MO, LO), PL, vHP, EPN, and STN, which was completely absent in saline-treated rats. It is also noticeable that WAY-163909 (3 mg/kg) increased the correlation between the striatal DOPAC/DA ratio with that of the frontal cortex, which was not seen in saline and WAY-163909 (0.3 mg/kg)-treated rats. Thus, WAY-163909 favored correlations of DOPAC/DA from diencephalic/mesencephalic regions toward other brain regions at 0.3 mg/kg and from cortical regions toward cortical and subcortical regions at 3 mg/kg.

2.1.3. Pattern of Correlations between DOPAC/DA and 5-HIAA/5-HT Ratio across the Brain

We looked at the correlations established between DA metabolism (DOPAC/DA ratio) and 5-HT metabolism (5-HIAA/5-HT ratio) across selected brain regions (Figure 4). The DOPAC/DA and 5-HIAA/5-HT ratios significantly correlated in 73 brain areas (comprising 47 positive and 26 negative correlations), representing again less than 10% of all analyses. It included 3 correlations in the same brain area (all positive) of saline-treated rats. The correlations included the NAc core and shell, EPN, and SN. Interestingly, the number of correlations decreased after 0.3 mg/kg and 3 mg/kg WAY-163909 injection (64 and 65 respectively, mostly positive correlations). The proportion of correlations in the same brain region was significantly more in 0.3 mg/kg WAY-163909 treated rats 7 (5 positive) compared to saline-treated rats. WAY-163909 completely diminished the correlation of the NAc shell DOPAC/DA ratio with the 5-HIAA/5-HT ratio in the striatum and SN. Likewise, WAY-163909 also suppressed the correlation between the DOPAC/DA ratio with the 5-HIAA/5-HT ratio of frontal

cortex in basal ganglia mesencephalon (EPN and STN), as well as the basal ganglia mesencephalon (EPN and STN) DOPAC/DA ratio with the orbitofrontal cortex (LO, DLO, and M2). The number of correlations dramatically increased in the CE and hypothalamus after WAY-163909 (0.3 mg/kg) treatment. WAY-163909 (3 mg/kg) potentiated the correlation of DOPAC/DA ratio from DLO and M2 with different brain areas such as the cortex, hippocampus, amygdala, and striatum (DMS, VMS, VCS), which are absent in saline and WAY-163909 (0.3 mg/kg) treated rats. WAY-163909 (0.3 and 3 mg/kg) also considerably potentiated the correlation of DA turnover with 5-HT turnover in MR, which is absent in saline-treated rats. Markedly, WAY-163909 at 0.3 mg/kg enhanced the correlations involving both the dHY and the vHY.

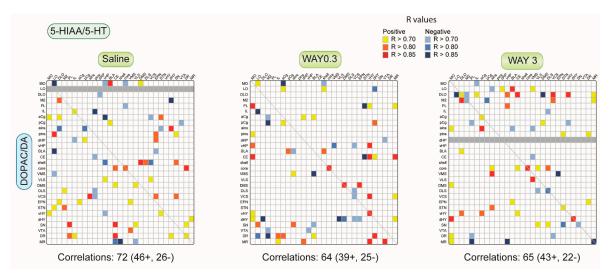


Figure 4. Correlative analysis between the 5-HT turnover (5-HIAA/5-HT ratio) and DA turnover (DOPAC/DA ratio) across rat brain regions. Representation of the range of Pearson's *r* values for each linear regression of 5-HIAA/5-HT ratio (top horizontal) versus DOPAC/DA ratio (left vertical) within and between the 29 brain areas in saline (first column), WAY-163909 0.3 mg/kg (second column) and 3 mg/kg (third column) treated rats. Colored boxes correspond to the existence of a significant correlation between the two parameters (yellow to red: positive; blue: negative) considered after correction for multiple comparisons. The grey boxes indicate that these correlations were not performed due to the low number of values (see Table S1).

3. Discussion

The results show that the 5-HT_{2C} receptor agonist WAY-163909 modifies the metabolism of 5-HT and DA systems in very few brain regions, with a noticeable increase in HVA content in the medial orbitofrontal cortex. Beyond quantitative, neurochemical changes, the results obtained with the correlations suggest that WAY-163909 rebalances the pattern of correlations of tissue monoamine metabolism in the brain.

The study of the metabolites and turnovers of 5-HT and DA systems brings up additional information on the impact of WAY-163909 on these systems, notably the increase in HVA or the ratio of DOPAC/DA in the MO cortex. Even if the purpose of our approach is not to speculate on the meaning of these results in terms of extracellular levels of DA, an increase in HVA in cortical regions is often interpreted as a consequence of an increase in DA release. Indeed, the clearance of extracellular DA is poorly achieved by DA reuptake sites, expressed at low levels in the orbitofrontal cortex. This strengthens the role of cortical, ectopic COMT in the degradation of DA [40]. The increase in HVA content parallels the increase in the ratio of DOPAC/DA in a situation where DA tissue content was not significantly altered [29]. It is important to stress that these effects are specific in this region. Moreover, we confirmed with the previous data that the quantitative, neurochemical changes for DA function are rather modest: increased DA in the CE at 0.3 mg/kg and decreased DA in the dHY [29], in addition to the decrease in DOPAC/DA turnover in ains at 0.3 mg/kg.

The changes are also modest regarding the metabolism of the 5-HT system, since we reported opposite changes of 5-HIAA tissue content in DLO and CE after 0.3 mg/kg and a decrease in the 5-HT turnover only in the IL. The level of 5-HT was previously shown to be reduced in DLO at 0.3 mg/kg and increased in MO and M2 at 3 mg/kg. Previous data reported no modification on tissue content of 5-HT and turnover in a restricted number of regions with the 5-HT_{2B/2C} receptor agonist Ro-60-0175 [41].

It can be argued that tissue measurement of monoamine metabolism is less sensitive to detecting changes of monoamine function compared to more dynamic *in vivo* methods. Indeed, it has been shown on many occasions that the stimulation of central 5-HT_{2C} receptors inhibited the electrical activity of the VTA DA neurons [42,43] and DA release in the nucleus accumbens and/or the striatum [44-46], while likely not in the medial prefrontal cortex [47,48]. Nonetheless, it is now acknowledged that several other sources of 5-HT_{2C} receptors can modulate DA release, sometimes in an opposite manner [23,49,50], which may explain the differences in DA output according to the agonists used [1]. WAY-163909, probably one of the most selective 5-HT_{2C} receptor agonists available [1,51,52], does not modulate DA release in the nucleus accumbens or striatum at 1 mg/kg nor modify c-Fos expression in the basal ganglia except in the ventrolateral striatum (VLS) at 3mg/kg [24]. The effects of WAY-163909 on DA release occurred at higher doses increased in the cortex and decreased in nucleus accumbens [48]. Thus, even if the approach is less sensitive, it allows the study of monoaminergic metabolism in multiple regions encompassing several neurobiological networks. The correlation changes for the 5-HIAA/5-HT and DOPAC/DA ratios support the idea that WAY-163909 should trigger widespread effects in the brain while being behaviorally efficacious with the regimen we used for purposeless oral movements, body temperature, probabilistic reversal learning in mice, and food intake, to cite a few [24,48,53–55].

The existing correlations of the tissue content for a monoamine or its metabolism between two separated brain regions are not easily interpreted. In fact, the regulation of biochemical activity at terminals includes autoregulations and heteroregulations at the vicinity, the latter ones being included into neurobiological networks. A single DA neuron from the SN crosses many distinct networks and establishes thousands of presynaptic interactions with other neurons [56]. Speculatively, it is most likely that the biochemical activity in one part of the monoaminergic neuron does not correlate with another part of the same neuron. The tissue measurement of monoamines and metabolites grossly integrates all homologous and heterologous regulating mechanisms within the piece of tissue. It is not surprising that the level of correlations is low at least in rats that underwent behavioral tests without receiving any injection [32,39]. The changes of correlations of a parameter between two brain regions would indicate a change of allegiance of the terminals toward regulatory mechanisms impacting the activity of the terminals, either homologous or heterologous or both, even in the absence of net changes of quantities. WAY-163909 favored cortico-cortical and cortico-subcortical correlations for DOPAC/DA at both doses and for 5-HIAA/5-HT at 0.3 mg/kg. The correlations are very different between saline, 0.3, and 3 mg/kg WAY-163909 treated rats and do not go over 10% of the total analyses. A completely different pattern was also triggered by WAY-163909 for the parent neurotransmitter, although the number of correlations was reduced for DA tissue content and enhanced for 5-HT and noradrenaline tissue contents [29]. Similarly, the reduction of correlations was small after WAY-163909 when comparing 5-HIAA/5-HT versus DOPAC/DA, but the profile was very different. At 3 mg/kg, the frontal DOPAC/DA ratios established several correlations with cortical and subcortical 5-HIAA/5-HT ratios. The distinct pattern of effects elicited by 0.3 and 3 mg/kg could have mainly two origins. As far as the 5-HT_{2C} receptors are concerned, it has to be considered that the 5-HT_{2C} receptor mRNA is post-transcriptionally edited at five distinct sites of its coding sequence [57]. Higher levels of editing reduce the affinity for ligands and the coupling efficiency for intracellular signaling pathways [58–60]. The profile of editing shows some regional specificity [61], as it could participate in the seemingly biphasic response induced by 0.3 and 3 mg/kg WAY-163909. The second origin for the reported differences could be an action of WAY-163909 on other targets. WAY-163909 has a 20-fold and 46-fold binding selectivity over the 5-HT_{2A} and 5-HT_{2B} receptors, respectively [51]. A progressive involvement of 5-HT_{2A} receptors for which it would behave as an antagonist [51] cannot be excluded.

differences which impact the DOPAC/DA and 5-HIAA/5-HT ratios in some brain regions [62], the precise location of the sampled brain regions from an experiment to another, the affective status of the animals, the systematic pooling of both sides of the brain for one brain region, and perhaps the number of animals in one group. In fact, we could not reproduce the correlations of the DOPAC/DA ratios between the striatal territories we previously reported on a large cohort of Wistar rats [39] or in adolescent male C57Bl6 mice [63]. In an almost similar experimental condition using male Sprague Dawley rats, we actually found some correlations of DOPAC/DA between striatal quadrants (including the shell and the core) and fewer negative correlations compared to the present study (unpublished observation). Although we do not have an explanation for these differences, the slightly higher number of animals (10 instead of 8 in the present study) could be an additional explanation as previously discussed [39].

It has been proposed that the 5-HT_{2C} receptor plays the role of neural rheostat shaping networks that overlap in behavioral dimensions, a hypothesis possibly explaining vulnerability to drug abuse [2]. Without strong quantitative effects of WAY-163909 on various electrophysiological, biochemical, or neuroanatomical markers except for the ones we report here, it is tempting to speculate that some changes of correlations induced by WAY-163909 could correspond to a remodeling of neurobiological networks, redefining overlapping networks as receptor fields. WAY-163909 lowered the sensitivity threshold of the basal ganglia to the stimulation of the cingulate cortex [24]. The magnitude of an existing response was not modified by WAY-163909 in contrast to the 5-HT_{2B/2C} receptor agonist Ro 60-0175 [64], and the effect of WAY-163909 on the threshold sensitivity was attributed to an action on the size of segregated circuits of the basal ganglia. This hypothesis deserves attention but would require adequate neuronal/tissue markers. The correlative analysis of monoamine tissue indexes is interesting because it highlights tissue actions which parallel behavioral effects of 5-HT_{2C} receptor agonists on the impulsive/compulsive dimension and cognitive flexibility [2,16,65–67], anxious states [68–71], or food intake [5,52,72,73]. Nevertheless, even if this approach has several dimensions with the study of the neurotransmitter (intraneuronal pool), metabolites, and turnovers, it is only descriptive and barely highlights the mechanisms of interaction of 5-HT_{2C} agonists in the brain.

In conclusion, the 5-HT_{2C} receptor agonist WAY-163909 modified the metabolites and the turnover indexes of 5-HT and DA only in a few brain regions. With an almost similar number of correlations of DOPAC/DA or 5-HIAA/5-HT between pairs of brain regions, WAY-163909 changed the whole pattern of correlations for both indexes. It is possible that the therapeutic actions of 5-HT_{2C} receptor agonists in diseases like obesity, abuse of drugs, or schizophrenia involves multiple changes over neurobiological networks rather than specific effects in one of them [1].

4. Materials and Methods

4.1. Animals

Male Sprague Dawley rats weighing 300–400 g were used (24 rats in total; Centre d'élevage R Janvier, Le Genest-Saint-Isle, France). They were kept in an animal facility (University of Bordeaux, France) with free access to food and water, in constant temperature ($21 \pm 2 \,^{\circ}$ C) and humidity (60%) levels, under a 12-h day/night cycle. All the animals' procedures were in accordance with the European Council Directive 2010/63/EU and the French National Committee (décret 2001-464) and local committee for the care and use of laboratory animals (N°50120130-A, 05/02/2013). All efforts were made to minimize animal suffering and to reduce the number of animals used.

4.2. Tissue Sampling and Conditioning

The whole procedure has been recently reported [29]. After the sacrifice, brains were frozen and cut using a cryostat at -24 °C. Bilateral "punches" were made of various brain structures of interest

using steel cannulae except for the subthalamic nucleus (STN), which was gently and progressively collected from the surface.

The selected brain regions belonged to different neurobiological networks and were selected with the help of a brain atlas [74]. Selected bilateral pieces of tissue for one brain region were placed in previously weighed Eppendorf tubes and brought to the freezer at -80 °C until the day of the biochemical analysis. Photos were taken throughout to ensure that the samples were taken from a similar plane each time. After the weighing of the filled tube to determine the weight of the tissue, 100 µL of perchloric acid (HClO₄ 0.1N, 4 °C) was added. The sonication of the samples preceded their centrifugation (13,000 rpm, 30 min, 4 °C), and part of the supernatant was injected into the high-pressure liquid chromatography (HPLC) system.

4.3. HPLC Analysis and Electrochemical Detection

The tissue concentrations of monoamines and metabolites were measured by HPLC coupled to the coulometric detection system using conditions previously reported [29]. The monoamines exited the column at different retention times (approximately DOPAC: 5'30; DA: 8'; 5-HIAA: 10'20"; HVA: 15'; 5-HT: 18'). They passed into the coulometric detection cell (Cell 5011, ESA, Paris, France), where the potential of the two electrodes was fixed via the coulometric detector (CoulochemII, ESA, Paris, France) at +350 mV (oxidation) and -270 mV (reduction), respectively. The coulometric detector was connected to a computer through an interface (Ulyss, Azur system, Toulouse, France).

The calibration curves were adapted to the brain areas investigated due to the heterogeneous distribution of monoamines and metabolites across the brain [32,39]. We also adapted a gains set at the level of the detector using a timeline method. The gains used ranged from 5 nA (for the DOPAC, DA, HVA in the hippocampus) to 1 μ A (for DA in the striatum). Standard solutions were used before each series of 10/12 samples to verify the good correspondence of the chromatographic conditions. The composition of the standards varied depending on the brain region investigated. The overall sensitivity for the compounds (in pg/10 μ L and its expression in fmol): DA, 2.3 (14.5 fmol); DOPAC, 3.4 (20.2); 5-HIAA, 5.5 (28.8); HVA, 18 (98.8); 5-HT, 7.6 (43.1) with a signal/noise ratio of 3:1.

We used two or even three HPLC systems in parallel, which could slightly differ in their sensitivity. In any case, all the samples for one brain region were analyzed with the same HPLC system.

4.4. Pharmacological Treatment and Experimental Design

WAY-163909 (7bR,10aR)-1,2, 3,4,8,9,10,10a-octahydro-7bH-cyclopenta-[b][1,4]diazepino[6,7, 1hi]indole]), a generous gift from Dr. John Dunlop (Wyeth Research, USA), was freshly diluted as free base in NaCl 0.9% the day of the experiment. It was injected i.p. (0.3 or 3 mg/kg). For each experimental group, the animals were randomized and received either the drug (0.3 or 3 mg/kg) or appropriate vehicle (in 1 ml/kg). The three treatments were equally distributed for the 24 rats (n = 8 for each treatment). The doses of 0.3 and 3 mg/kg, i.p. were chosen based on previous data showing that they reduced locomotor activity and induced purposeless oral movements [48,75]. These studies reported maximal behavioral effects after 30 minutes, and we correspondingly stopped the experiment 45 minutes after its injection at a time corresponding to the highest behavioral activity. We also briefly evaluated abnormal oral movements and penile grooming without scoring them in this experiment.

4.5. Statistical Analysis of the Data

The tissue levels of 5-HT and DA for each structure have been reported in a previous article [29]. Here, we report the values expressed in pg/mg for 5-HIAA, DOPAC, and HVA in each brain region. These levels are presented as the mean \pm the standard error of the mean (SEM) according to their treatment group. Outlier data were discarded on the basis of the value outside the range of the average mean \pm two standard deviations [39], though there were few detected outliers. On the other hand, data were lost due to accidental manipulations (e.g., piece of tissue lost during the punching procedure, aberrant determination of the weight of the piece of tissue). Additionally, several data

were lost for HVA due to the poor sensitivity and/or low levels, and groups having lost half of the data (4 observations on 8) were discarded. We calculated the index of the 5-HT and DA turnovers as 5-HIAA/5-HT and DOPAC/DA for each animal and structure and reported the values as mean \pm SEM. The values for 5-HIAA, DOPAC, HVA, 5-HIAA/5-HT, and DOPAC/DA were compared between experimental groups (saline, WAY-163909 at 0.3 or 3 mg/kg, i.p.) with a one-way ANOVA, followed by the Fisher protected least significant difference (PLSD) post-hoc test. *p* < 0.05 was used as the criterion for significance.

The qualitative analysis corresponds to multiple correlative analyses using Bravais–Pearson's correlation coefficient. These analyzes were performed within and between 5-HIAA/5-HT and DOPAC/DA in the 29 brain regions investigated. The correlations were separately performed in rats receiving saline, 0.3 mg/kg WAY-163909, and 3 mg/kg WAY-163909 to estimate connectivity. As previously reported [32], *p*-values were adjusted using the false discovering rate (FDR) controlling procedures. Correlations were then considered as significant at the 5% level and were reported in the corresponding figures. Statistics were done using SigmaPlot 11.0 software.

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Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

| aCg | anterior cingulate cortex |
|--------|-------------------------------------|
| ains | anterior insular cortex |
| BLA | basolateral nucleus of the amygdala |
| CE | central nucleus of the amygdala |
| CNS | central nervous system |
| core | core of the nucleus accumbens |
| DA | dopamine |
| dHP | dorsal hippocampus |
| DLO | dorsolateral orbitofrontal cortex |
| DLS | dorsolateral striatum |
| DMS | dorsomedial striatum |
| DOPAC | 3,4-dihydroxyphenylacetic acid |
| DR | dorsal raphe nucleus |
| dHY | dorsal hypothalamus |
| EPN | entodepuncular nucleus |
| HPLC | high pressure liquid chromatography |
| IL | infralimbic cortex |
| LO | lateral orbitofrontal cortex |
| M2 | motor cortex M2 |
| MO | medial orbitofrontal cortex |
| MR | median raphe nucleus |
| pCg | posterior cingulate cortex |
| pins | posterior insular cortex |
| PL | prelimbic cortex |
| 5-HIAA | 5-hydroxyindole-3-acetic acid |
| 5-HT | 5-hydroxytryptamine; serotonin |
| | |

| $5-HT_{2C}$ receptors | Serotonin 2C receptors |
|-----------------------|---|
| shell | shell of the nucleus accumbens |
| SN | substantia nigra |
| STN | subthalamic nucleus |
| vHP | ventral hippocampus |
| vHY | ventral hypothalamus |
| VCS | ventrocaudal striatum |
| VLS | ventrolateral striatum |
| VMS | ventromedial striatum |
| VTA | ventral tegmental area |
| WAY-163909 | (7bR,10aR)-1,2,3,4,8,9,10,10a-octahydro-7bH-cyclopenta-[b][1,4]diazepino[6,7, 1hi]indole] |
| | |

References

- 1. Di Giovanni, G.; De Deurwaerdere, P. New therapeutic opportunities for 5-ht2c receptor ligands in neuropsychiatric disorders. *Pharmacol. Ther.* **2016**, *157*, 125–162. [CrossRef] [PubMed]
- Anastasio, N.C.; Stutz, S.J.; Fox, R.G.; Sears, R.M.; Emeson, R.B.; DiLeone, R.J.; O'Neil, R.T.; Fink, L.H.; Li, D.; Green, T.A.; et al. Functional status of the serotonin 5-ht2c receptor (5-ht2cr) drives interlocked phenotypes that precipitate relapse-like behaviors in cocaine dependence. *Neuropsychopharmacology* 2014, *39*, 370–382. [CrossRef] [PubMed]
- 3. Tecott, L.H.; Sun, L.M.; Akana, S.F.; Strack, A.M.; Lowenstein, D.H.; Dallman, M.F.; Julius, D. Eating disorder and epilepsy in mice lacking 5-ht2c serotonin receptors. *Nature* **1995**, *374*, 542–546. [CrossRef] [PubMed]
- D'Agostino, G.; Lyons, D.; Cristiano, C.; Lettieri, M.; Olarte-Sanchez, C.; Burke, L.K.; Greenwald-Yarnell, M.; Cansell, C.; Doslikova, B.; Georgescu, T.; et al. Nucleus of the solitary tract serotonin 5-ht2c receptors modulate food intake. *Cell Metab.* 2018, 28, 619–630. [CrossRef] [PubMed]
- 5. Clifton, P.G.; Lee, M.D.; Dourish, C.T. Similarities in the action of ro 60-0175, a 5-ht2c receptor agonist and d-fenfluramine on feeding patterns in the rat. *Psychopharmacology* **2000**, *152*, 256–267. [CrossRef] [PubMed]
- 6. Higgins, G.A.; Fletcher, P.J. Serotonin and drug reward: Focus on 5-ht2c receptors. *Eur. J. Pharmacol.* **2003**, *480*, 151–162. [CrossRef] [PubMed]
- 7. Higgins, G.A.; Fletcher, P.J. Therapeutic potential of 5-ht2c receptor agonists for addictive disorders. *ACS Chem. Neurosci.* 2015, *6*, 1071–1088. [CrossRef] [PubMed]
- 8. Howell, L.L.; Cunningham, K.A. Serotonin 5-ht2 receptor interactions with dopamine function: Implications for therapeutics in cocaine use disorder. *Pharmacol. Rev.* **2015**, *67*, 176–197. [CrossRef]
- 9. Fletcher, P.J.; Soko, A.D.; Higgins, G.A. Impulsive action in the 5-choice serial reaction time test in 5-ht(2)c receptor null mutant mice. *Psychopharmacology* **2013**, *226*, 561–570. [CrossRef]
- Anastasio, N.C.; Stutz, S.J.; Fink, L.H.; Swinford-Jackson, S.E.; Sears, R.M.; DiLeone, R.J.; Rice, K.C.; Moeller, F.G.; Cunningham, K.A. Serotonin (5-ht) 5-ht2a receptor (5-ht2ar):5-ht2cr imbalance in medial prefrontal cortex associates with motor impulsivity. ACS Chem. Neurosci. 2015, 6, 1248–1258. [CrossRef]
- Siuciak, J.A.; Chapin, D.S.; McCarthy, S.A.; Guanowsky, V.; Brown, J.; Chiang, P.; Marala, R.; Patterson, T.; Seymour, P.A.; Swick, A.; et al. Cp-809,101, a selective 5-ht2c agonist, shows activity in animal models of antipsychotic activity. *Neuropharmacology* 2007, *52*, 279–290. [CrossRef] [PubMed]
- 12. Kennett, G.A.; Whitton, P.; Shah, K.; Curzon, G. Anxiogenic-like effects of mcpp and tfmpp in animal models are opposed by 5-htlc receptor antagonists. *Eur. J. Pharmacol.* **1989**, *164*, 445–454. [CrossRef]
- 13. Millan, M.J. The neurobiology and control of anxious states. Prog. Neurobiol. 2003, 70, 83–244. [CrossRef]
- 14. Millan, M.J. Serotonin 5-ht2c receptors as a target for the treatment of depressive and anxious states: Focus on novel therapeutic strategies. *Therapie* **2005**, *60*, 441–460. [CrossRef] [PubMed]
- 15. Graf, M. 5-ht2c receptor activation induces grooming behaviour in rats: Possible correlations with obsessive-compulsive disorder. *Off. J. Hung. Assoc. Psychopharmacol.* **2006**, *8*, 23–28.
- 16. Kreiss, D.S.; De Deurwaerdere, P. Purposeless oral activity induced by meta-chlorophenylpiperazine (m-cpp): Undefined tic-like behaviors? *J. Neurosci. Methods* **2017**, *292*, 30–36. [CrossRef]
- 17. De Gregorio, D.; Enns, J.P.; Nunez, N.A.; Posa, L.; Gobbi, G. D-lysergic acid diethylamide, psilocybin, and other classic hallucinogens: Mechanism of action and potential therapeutic applications in mood disorders. *Prog. Brain Res.* **2018**, *242*, 69–96.

- 18. Chagraoui, A.; Thibaut, F.; Skiba, M.; Thuillez, C.; Bourin, M. 5-ht2c receptors in psychiatric disorders: A review. *Prog. Neuro-Psychopharmacol. Biol. Psychiatr.* **2016**, *66*, 120–135. [CrossRef]
- Abramowski, D.; Rigo, M.; Duc, D.; Hoyer, D.; Staufenbiel, M. Localization of the 5-hydroxytryptamine2c receptor protein in human and rat brain using specific antisera. *Neuropharmacology* 1995, 34, 1635–1645. [CrossRef]
- 20. Clemett, D.A.; Punhani, T.; Duxon, M.S.; Blackburn, T.P.; Fone, K.C. Immunohistochemical localisation of the 5-ht2c receptor protein in the rat cns. *Neuropharmacology* **2000**, *39*, 123–132. [CrossRef]
- Pasqualetti, M.; Ori, M.; Castagna, M.; Marazziti, D.; Cassano, G.B.; Nardi, I. Distribution and cellular localization of the serotonin type 2c receptor messenger rna in human brain. *Neuroscience* 1999, 92, 601–611. [CrossRef]
- 22. Pazos, A.; Palacios, J.M. Quantitative autoradiographic mapping of serotonin receptors in the rat brain. I. Serotonin-1 receptors. *Brain Res.* **1985**, *346*, 205–230. [CrossRef]
- De Deurwaerdere, P.; Lagiere, M.; Bosc, M.; Navailles, S. Multiple controls exerted by 5-ht2c receptors upon basal ganglia function: From physiology to pathophysiology. *Exp. Brain Res.* 2013, 230, 477–511. [CrossRef] [PubMed]
- 24. Lagiere, M.; Bosc, M.; Whitestone, S.; Manem, J.; Elboukhari, H.; Benazzouz, A.; Di Giovanni, G.; De Deurwaerdere, P. Does the serotonin2c receptor segregate circuits of the basal ganglia responding to cingulate cortex stimulation? *CNS Neurosci. Ther.* **2018**, *24*, 741–744. [CrossRef] [PubMed]
- 25. Navailles, S.; Lagiere, M.; Le Moine, C.; De Deurwaerdere, P. Role of 5-ht2c receptors in the enhancement of c-fos expression induced by a 5-ht2b/2c inverse agonist and 5-ht 2 agonists in the rat basal ganglia. *Exp. Brain Res.* **2013**, *230*, 525–535. [CrossRef] [PubMed]
- 26. De Deurwaerdere, P.; Di Giovanni, G. Serotonergic modulation of the activity of mesencephalic dopaminergic systems: Therapeutic implications. *Prog. Neurobiol.* **2017**, *151*, 175–236. [CrossRef] [PubMed]
- 27. Eisenhofer, G.; Kopin, I.J.; Goldstein, D.S. Catecholamine metabolism: A contemporary view with implications for physiology and medicine. *Pharmacol. Rev.* **2004**, *56*, 331–349. [CrossRef] [PubMed]
- 28. Chesselet, M.F. Presynaptic regulation of neurotransmitter release in the brain: Facts and hypothesis. *Neuroscience* **1984**, *12*, 347–375. [CrossRef]
- Chagraoui, A.; Whitestone, S.; Baassiri, L.; Manem, J.; Di Giovanni, G.; De Deurwaerdere, P. Neurochemical impact of the 5-ht2c receptor agonist way-163909 on monoamine tissue content in the rat brain. *Neurochem. Int.* 2019, 124, 245–255. [CrossRef]
- 30. Commissiong, J.W. Monoamine metabolites: Their relationship and lack of relationship to monoaminergic neuronal activity. *Biochem. Pharmacol.* **1985**, *34*, 1127–1131. [CrossRef]
- 31. Sharp, T.; Zetterstrom, T.; Ungerstedt, U. An in vivo study of dopamine release and metabolism in rat brain regions using intracerebral dialysis. *J. Neurochem.* **1986**, *47*, 113–122. [CrossRef] [PubMed]
- 32. Fitoussi, A.; Dellu-Hagedorn, F.; De Deurwaerdere, P. Monoamines tissue content analysis reveals restricted and site-specific correlations in brain regions involved in cognition. *Neuroscience* **2013**, 255, 233–245. [CrossRef] [PubMed]
- Navailles, S.; Bioulac, B.; Gross, C.; De Deurwaerdere, P. Chronic l-dopa therapy alters central serotonergic function and l-dopa-induced dopamine release in a region-dependent manner in a rat model of parkinson's disease. *Neurobiol. Dis.* 2011, 41, 585–590. [CrossRef] [PubMed]
- 34. Cooper, J.R.; Bloom, F.E.; Roth, R.H. *The Biochemical Basis of Neuropharmacology*, 8th ed.; Oxford University Press: New York, NY, USA, 2003.
- 35. De Deurwaerdere, P.; Binda, C.; Corne, R.; Leone, C.; Valeri, A.; Valoti, M.; Ramsay, R.R.; Fall, Y.; Marco-Contelles, J. Comparative analysis of the neurochemical profile and mao inhibition properties of n-(furan-2-ylmethyl)-n-methylprop-2-yn-1-amine. *ACS Chem. Neurosci.* **2017**, *8*, 1026–1035. [CrossRef] [PubMed]
- Bortolato, M.; Godar, S.C.; Alzghoul, L.; Zhang, J.; Darling, R.D.; Simpson, K.L.; Bini, V.; Chen, K.; Wellman, C.L.; Lin, R.C.; et al. Monoamine oxidase a and a/b knockout mice display autistic-like features. *Int. J. Neuropsychopharmacol.* 2013, *16*, 869–888. [CrossRef]
- 37. Shih, J.C.; Chen, K. Mao-a and -b gene knock-out mice exhibit distinctly different behavior. *Neurobiology* **1999**, *7*, 235–246.
- 38. De Deurwaerdere, P.; Di Giovanni, G.; Millan, M.J. Expanding the repertoire of l-dopa's actions: A comprehensive review of its functional neurochemistry. *Prog. Neurobiol.* **2017**, *151*, 57–100. [CrossRef]

- Dellu-Hagedorn, F.; Fitoussi, A.; De Deurwaerdere, P. Correlative analysis of dopaminergic and serotonergic metabolism across the brain to study monoaminergic function and interaction. *J. Neurosci. Methods* 2017, 280, 54–63. [CrossRef]
- 40. Kaenmaki, M.; Tammimaki, A.; Myohanen, T.; Pakarinen, K.; Amberg, C.; Karayiorgou, M.; Gogos, J.A.; Mannisto, P.T. Quantitative role of comt in dopamine clearance in the prefrontal cortex of freely moving mice. *J. Neurochem.* **2010**, *114*, 1745–1755. [CrossRef]
- 41. Mongeau, R.; Martin, C.B.; Chevarin, C.; Maldonado, R.; Hamon, M.; Robledo, P.; Lanfumey, L. 5-ht2c receptor activation prevents stress-induced enhancement of brain 5-ht turnover and extracellular levels in the mouse brain: Modulation by chronic paroxetine treatment. *J. Neurochem.* **2010**, *115*, 438–449. [CrossRef]
- 42. Di Giovanni, G.; Di Matteo, V.; Di Mascio, M.; Esposito, E. Preferential modulation of mesolimbic vs. Nigrostriatal dopaminergic function by serotonin(2c/2b) receptor agonists: A combined in vivo electrophysiological and microdialysis study. *Synapse* **2000**, *35*, 53–61. [CrossRef]
- Di Matteo, V.; Di Giovanni, G.; Di Mascio, M.; Esposito, E. Biochemical and electrophysiological evidence that ro 60-0175 inhibits mesolimbic dopaminergic function through serotonin(2c) receptors. *Brain Res.* 2000, *865*, 85–90. [CrossRef]
- De Deurwaerdere, P.; Navailles, S.; Berg, K.A.; Clarke, W.P.; Spampinato, U. Constitutive activity of the serotonin2c receptor inhibits in vivo dopamine release in the rat striatum and nucleus accumbens. *J. Neurosci.* 2004, 24, 3235–3241. [CrossRef] [PubMed]
- 45. Gobert, A.; Rivet, J.M.; Lejeune, F.; Newman-Tancredi, A.; Adhumeau-Auclair, A.; Nicolas, J.P.; Cistarelli, L.; Melon, C.; Millan, M.J. Serotonin(2c) receptors tonically suppress the activity of mesocortical dopaminergic and adrenergic, but not serotonergic, pathways: A combined dialysis and electrophysiological analysis in the rat. *Synapse* **2000**, *36*, 205–221. [CrossRef]
- Browne, C.J.; Ji, X.; Higgins, G.A.; Fletcher, P.J.; Harvey-Lewis, C. Pharmacological modulation of 5-ht2c receptor activity produces bidirectional changes in locomotor activity, responding for a conditioned reinforcer, and mesolimbic da release in c57bl/6 mice. *Neuropsychopharmacology* 2017, 42, 2178–2187. [CrossRef] [PubMed]
- Pozzi, L.; Acconcia, S.; Ceglia, I.; Invernizzi, R.W.; Samanin, R. Stimulation of 5-hydroxytryptamine (5-ht(2c)) receptors in the ventrotegmental area inhibits stress-induced but not basal dopamine release in the rat prefrontal cortex. *J. Neurochem.* 2002, *82*, 93–100. [CrossRef] [PubMed]
- Marquis, K.L.; Sabb, A.L.; Logue, S.F.; Brennan, J.A.; Piesla, M.J.; Comery, T.A.; Grauer, S.M.; Ashby, C.R., Jr.; Nguyen, H.Q.; Dawson, L.A.; et al. Way-163909 [(7br,10ar)-1,2,3,4,8,9,10,10a-octahydro-7bh-cyclopenta-[b][1,4]diazepino[6,7,1hi]indole]: A novel 5-hydroxytryptamine 2c receptor-selective agonist with preclinical antipsychotic-like activity. *J. Pharmacol. Exp. Ther.* 2007, 320, 486–496. [CrossRef] [PubMed]
- 49. Lucas, G.; Spampinato, U. Role of striatal serotonin2a and serotonin2c receptor subtypes in the control of in vivo dopamine outflow in the rat striatum. *J. Neurochem.* **2000**, *74*, 693–701. [CrossRef] [PubMed]
- 50. Leggio, G.M.; Cathala, A.; Neny, M.; Rouge-Pont, F.; Drago, F.; Piazza, P.V.; Spampinato, U. In vivo evidence that constitutive activity of serotonin2c receptors in the medial prefrontal cortex participates in the control of dopamine release in the rat nucleus accumbens: Differential effects of inverse agonist versus antagonist. *J. Neurochem.* 2009, 111, 614–623. [CrossRef]
- Dunlop, J.; Marquis, K.L.; Lim, H.K.; Leung, L.; Kao, J.; Cheesman, C.; Rosenzweig-Lipson, S. Pharmacological profile of the 5-ht(2c) receptor agonist way-163909; therapeutic potential in multiple indications. CNS Drug Rev. 2006, 12, 167–177. [CrossRef]
- Dunlop, J.; Sabb, A.L.; Mazandarani, H.; Zhang, J.; Kalgaonker, S.; Shukhina, E.; Sukoff, S.; Vogel, R.L.; Stack, G.; Schechter, L.; et al. Way-163909 [(7br, 10ar)-1,2,3,4,8,9,10,10a-octahydro-7bh-cyclopenta-[b][1,4]diazepino[6,7,1hi]indol e], a novel 5-hydroxytryptamine 2c receptor-selective agonist with anorectic activity. *J. Pharmacol. Exp. Ther.* 2005, *313*, 862–869. [CrossRef] [PubMed]
- 53. Murphy, T.J.; Murnane, K.S. The serotonin 2c receptor agonist way-163909 attenuates ketamine-induced hypothermia in mice. *Eur. J. Pharmacol.* **2019**, *8*42, 255–261. [CrossRef] [PubMed]
- 54. Phillips, B.U.; Dewan, S.; Nilsson, S.R.O.; Robbins, T.W.; Heath, C.J.; Saksida, L.M.; Bussey, T.J.; Alsio, J. Selective effects of 5-ht2c receptor modulation on performance of a novel valence-probe visual discrimination task and probabilistic reversal learning in mice. *Psychopharmacology* 2018, 235, 2101–2111. [CrossRef] [PubMed]

- 55. Berro, L.F.; Perez Diaz, M.; Maltbie, E.; Howell, L.L. Effects of the serotonin 2c receptor agonist way163909 on the abuse-related effects and mesolimbic dopamine neurochemistry induced by abused stimulants in rhesus monkeys. *Psychopharmacology* **2017**, *234*, 2607–2617. [CrossRef] [PubMed]
- 56. Matsuda, W.; Furuta, T.; Nakamura, K.C.; Hioki, H.; Fujiyama, F.; Arai, R.; Kaneko, T. Single nigrostriatal dopaminergic neurons form widely spread and highly dense axonal arborizations in the neostriatum. *J. Neurosci.* **2009**, *29*, 444–453. [CrossRef]
- 57. Gardiner, K.; Du, Y. A-to-i editing of the 5ht2c receptor and behaviour. *Brief. Funct. Genom. Proteom.* **2006**, *5*, 37–42. [CrossRef]
- Berg, K.A.; Cropper, J.D.; Niswender, C.M.; Sanders-Bush, E.; Emeson, R.B.; Clarke, W.P. Rna-editing of the 5-ht(2c) receptor alters agonist-receptor-effector coupling specificity. *Br. J. Pharmacol.* 2001, 134, 386–392. [CrossRef]
- 59. Martin, C.B.; Ramond, F.; Farrington, D.T.; Aguiar, A.S., Jr.; Chevarin, C.; Berthiau, A.S.; Caussanel, S.; Lanfumey, L.; Herrick-Davis, K.; Hamon, M.; et al. Rna splicing and editing modulation of 5-ht(2c) receptor function: Relevance to anxiety and aggression in vgv mice. *Mol. Psychiatry* **2013**, *18*, 656–665. [CrossRef]
- 60. Niswender, C.M.; Sanders-Bush, E.; Emeson, R.B. Identification and characterization of rna editing events within the 5-ht2c receptor. *Ann. N. Y. Acad. Sci.* **1998**, *861*, 38–48. [CrossRef]
- 61. Burns, C.M.; Chu, H.; Rueter, S.M.; Hutchinson, L.K.; Canton, H.; Sanders-Bush, E.; Emeson, R.B. Regulation of serotonin-2c receptor g-protein coupling by rna editing. *Nature* **1997**, *387*, 303–308. [CrossRef]
- 62. Dellu-Hagedorn, F.; Rivalan, M.; Fitoussi, A.; De Deurwaerdere, P. Inter-individual differences in the impulsive/compulsive dimension: Deciphering related dopaminergic and serotonergic metabolisms at rest. *Philos. Trans. R. Soc. Lond. Ser. B Biol. Sci.* **2018**, *373.* [CrossRef] [PubMed]
- 63. Puginier, E.; Bharatiya, R.; Chagraoui, A.; Manem, J.; Cho, Y.H.; Garret, M.; De Deurwaerdere, P. Early neurochemical modifications of monoaminergic systems in the r6/1 mouse model of huntington's disease. *Neurochem. Int.* **2019**, *128*, 186–195. [CrossRef] [PubMed]
- 64. Beyeler, A.; Kadiri, N.; Navailles, S.; Boujema, M.B.; Gonon, F.; Moine, C.L.; Gross, C.; De Deurwaerdere, P. Stimulation of serotonin2c receptors elicits abnormal oral movements by acting on pathways other than the sensorimotor one in the rat basal ganglia. *Neuroscience* **2010**, *169*, 158–170. [CrossRef] [PubMed]
- Alsio, J.; Nilsson, S.R.; Gastambide, F.; Wang, R.A.; Dam, S.A.; Mar, A.C.; Tricklebank, M.; Robbins, T.W. The role of 5-ht2c receptors in touchscreen visual reversal learning in the rat: A cross-site study. *Psychopharmacology* 2015, 232, 4017–4031. [CrossRef] [PubMed]
- Flaisher-Grinberg, S.; Klavir, O.; Joel, D. The role of 5-ht2a and 5-ht2c receptors in the signal attenuation rat model of obsessive-compulsive disorder. *Int. J. Neuropsychopharmacol.* 2008, *11*, 811–825. [CrossRef] [PubMed]
- 67. Fletcher, P.J.; Rizos, Z.; Noble, K.; Higgins, G.A. Impulsive action induced by amphetamine, cocaine and mk801 is reduced by 5-ht(2c) receptor stimulation and 5-ht(2a) receptor blockade. *Neuropharmacology* **2011**, *61*, 468–477. [CrossRef] [PubMed]
- Bagdy, G.; Graf, M.; Anheuer, Z.E.; Modos, E.A.; Kantor, S. Anxiety-like effects induced by acute fluoxetine, sertraline or m-cpp treatment are reversed by pretreatment with the 5-ht2c receptor antagonist sb-242084 but not the 5-ht1a receptor antagonist way-100635. *Int. J. Neuropsychopharmacol.* 2001, *4*, 399–408. [CrossRef] [PubMed]
- 69. Campbell, B.M.; Merchant, K.M. Serotonin 2c receptors within the basolateral amygdala induce acute fear-like responses in an open-field environment. *Brain Res.* **2003**, *993*, 1–9. [CrossRef]
- 70. Jenck, F.; Bos, M.; Wichmann, J.; Stadler, H.; Martin, J.R.; Moreau, J.L. The role of 5-ht2c receptors in affective disorders. *Expert Opin. Investig. Drugs* **1998**, *7*, 1587–1599. [CrossRef]
- 71. Pockros-Burgess, L.A.; Pentkowski, N.S.; Der-Ghazarian, T.; Neisewander, J.L. Effects of the 5-ht2c receptor agonist cp809101 in the amygdala on reinstatement of cocaine-seeking behavior and anxiety-like behavior. *Int. J. Neuropsychopharmacol.* **2014**, *17*, 1751–1762. [CrossRef]
- 72. Hayashi, A.; Sonoda, R.; Kimura, Y.; Takasu, T.; Suzuki, M.; Sasamata, M.; Miyata, K. Antiobesity effect of ym348, a novel 5-ht2c receptor agonist, in zucker rats. *Brain Res.* **2004**, *1011*, 221–227. [CrossRef] [PubMed]
- 73. Rosenzweig-Lipson, S.; Zhang, J.; Mazandarani, H.; Harrison, B.L.; Sabb, A.; Sabalski, J.; Stack, G.; Welmaker, G.; Barrett, J.E.; Dunlop, J. Antiobesity-like effects of the 5-ht2c receptor agonist way-161503. *Brain Res.* 2006, 1073–1074, 240–251. [CrossRef] [PubMed]

- 74. Paxinos, G.; Watson, C. *The Rat Brain in Stereotaxic Coordinates*, 4th ed.; Academic Press: San Diego, CA, USA, 1998.
- 75. Navailles, S.; Lagiere, M.; Roumegous, A.; Polito, M.; Boujema, M.B.; Cador, M.; Dunlop, J.; Chesselet, M.F.; Millan, M.J.; De Deurwaerdere, P. Serotonin2c ligands exhibiting full negative and positive intrinsic activity elicit purposeless oral movements in rats: Distinct effects of agonists and inverse agonists in a rat model of parkinson's disease. *Int. J. Neuropsychopharmacol.* **2013**, *16*, 593–606. [CrossRef] [PubMed]



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